





### Application of Tailored Magnetic Nanoparticles for Protein Purification

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#### Introduction to magnetic nanoparticles



- Nanoparticles (<25 nm) of ferro-/ferrimagnetic materials show superparamagnetic behavior → no coercivity nor remanence
- This prevents an agglomeration by magnetic attractive forces
- Under an external field, high saturation magnetization is achieved
  → magnetic fluids (small, stable nanoparticles, high-η medium)
  - $\rightarrow$  magnetic nanoparticle manipulation (larger nanoparticles, low- $\eta$  medium)



### Aims and approach



- Highly efficient, selective and low-cost magnetic separation system for the purification of 6-Histidine tagged recombinant proteins
- Selective separation of target protein in cell-free supernatant (*ex situ*) / during cultivation in a bioreactor (*in situ*)
- Tailor-made <u>superparamagnetic iron oxide nanoparticles</u> (SPIONs)
- Great simplification of downstream processing by reduction of processing steps
- Recyclable system for cost efficiency



## **Synthesis of SPIONs**











### **Synthesis of SPIONs**



Non-aqueous sol-gel synthesis in triethylene glycol (TEG)



- Formation of highly crystalline nanoparticles
- Synthesis in 1 L reactor established
- Spherical SPIONs of 8 nm with good homogeneity obtained
- Size and composition can be altered for tailoring of magnetic properties



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# **Synthesis of SPIONs**





Analysis of particle formation and growth via kinetic studies in reactor system

- Formation via sol-gel-type process
- Organic content of particles diminishes over time
- Process conditions allow tuning of particle size to some extent
- Stable particle dispersions are obtained in TEG and H<sub>2</sub>O

























Coupling of (3-glycidyloxypropyl)trimethoxysilane (GLYMO) with N,N-bis(carboxymethyl)-L-lysine (NTA)

















- Model system: Recombinant Protein A from Bacillus megaterium cultures
- Selective separation of target protein in cell-free supernatant (*ex situ*) / during cultivation in a bioreactor (*in situ*)
- No adverse effects on bacterial productivity observed
- Use of SPIONs in multiple cycles





Scanning electron microscopy comparison of SPIONs and bacteria









In situ binding and separation efficiency

Constant SPION concentration of 500  $\mu$ g/mL culture 100 % = 60 mg Protein A per liter (120 mg/g SPIONs)

Product bound from media



- Purity was determined using SDS-PAGE with a sensitivity of 5 ng per sample
- No protein impurity in the elution fraction
- Final purity of <u>~ 99.9 %</u> regarding protein compounds.





Dynamic light scattering (DLS) measurements of particles after recycling



- The size of the regenerated particles does not differ significantly between the cycles.
- Particles are present as defined agglomerates of ~ 1000 nm in diameter.





# **Purification and regeneration – challenges**

- High separation efficiency can only be ensured when constant particle concentration is realized
- Loss of nanoparticles in each production and regeneration cycle
- Insufficient magnetic separation / incomplete precipitation due to varying colloidal stability



## Summary



#### **SPIONs with tailored properties**

- Non-aqueous synthesis as versatile and scalable strategy
- Process knowledge allows adjustment of properties
- Post-structuring into hierarchical aggregates

#### **Functionalization strategy**

- Use of designed linker allows covalent coupling
- Optimization of surface concentration
- Robust protocol for linker synthesis and coupling

#### **Purification of recombinant proteins**

- Metal complex-based protein binding strategy
- Purification and regeneration cycle established
- In situ purification successfully accomplished with high efficiency and purity



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